Why Despite Rumen Degradation of Mycotoxins Ruminants are Still at **Potential Risk for Mycotoxin Toxicity**

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Abstract

Despite widespread occurrence of mycotoxin contamination of ruminant diets is evident globally; ruminants were traditionally considered less susceptible to mycotoxins due to rumen microbial degradation. But, growing body of evidence is highlighting that tolerance of ruminants to mycotoxins is a 'consideration' but not a fool-proof process of complete mycotoxin-inactivation. Yet, remarkably there is a paucity of research studies of mycotoxins in ruminants than in monogastric farm animals (pigs and poultry). Available data show a low number of studies that are mainly related to feed intake, nutrient utilization/digestibility or on production performance (milk yield and growth rate). *In-vitro* studies mostly focus on rumen incubation and immune cells proliferation; with very few articles focusing on reproductive function. However, none of the articles have explicitly explained the diverse set of mechanistic link for the occurrence of deaths and many adverse effects being reported in ruminants following mycotoxin exposure. Thus in the present review, we summarizes the factual evidence explaining how despite rumen microbial action the mycotoxins are potential risk factors in ruminants diets. The current article further enlists and explains various factors that are contributing to the adverse effects of mycotoxins in ruminants despite mycotoxin level being below permissible regulatory limits.

Keywords: Mycotoxins, Ruminants, Animal feed, Adverse effects, Rumen microbes

YCOTOXINS are a global animal public health **W** concern including ruminants. Mycotoxins are secondary fungal metabolites with a worldwide spectrum of occurrence and consequently their exposure both in humans as well as animals is high globally. Alarmingly, the global occurrence of mycotoxins in animal feed is not new and has been a persistent scenario since several decades (Rodrigues, 2014; Rodrigues & Naehrer, 2012 and Biomin, 2020). A study performed in 2013 on feed samples from across the world demonstrated that approximately 70 per cent of samples tested was mycotoxin contaminated (Streit et al., 2013). It is no surprise that co-occurrence of multiple mycotoxins is quite common (Biomin, 2020). The situation is further complicated by increasing prevalence of mycotoxin

due to climate change (Battilani et al., 2016), putting the animals including ruminants at higher risk for mycotoxin exposure and related toxicity.

Mycotoxins in animal feed occur when fungi infects the food-crops during growth and harvesting (field mycotoxins) as well as when it infects the food-stuff during drying, transport and storage (storage mycotoxins). While corn, wheat, rice, barley, oats and rye are the common cereals affected by mycotoxins, nevertheless straw and silages are also contaminated. The most commonly occurring mycotoxins are produced mainly by the fungi of the genera, Aspergillus, Penicillium, Fusarium, Claviceps and Stachybotrys (Robbins et al., 2000). Although several hundred mycotoxins have been identified, the most common mycotoxins found to be implicated in their toxicities include aflatoxins, trichothecenes (T-2, deoxynivalenol - DON), fumonisins (FUM), zearalenone, ochratoxin and ergot alkaloids (Grenier and Oswald, 2011). Among them, fusarium mycotoxins (FUM, DON) were the most prevalent mycotoxins in animal feed in Europe, Asia and worldwide as well indicated by global survey reports (Rodrigues & Naehrer, 2012 and Biomin, 2020). In addition, more recently, there are emerging mycotoxins that are more-and-more detected in animal feed that includes enniatin, phomopsin A, alternariol, etc., suggesting that mycotoxins in ruminants is a critical issue due to their constant presence.

The problem with mycotoxins is their toxicity (mycotoxicosis) and associated disease. Deaths due to mycotoxin contamination are frequently reported both in humans as well as animals, such as death of 125 people in Kenya in 2004 and death of dogs in USA in 2006, following ingestion of aflatoxincontaminated food and feed, respectively (Lewis et al., 2005 and Susan S. Lang, 2006). More recently, over 110 dogs died following exposure to aflatoxins in pet food (FDA, 2021). However, it was previously considered that ruminants were less susceptible to mycotoxins due to degradation of mycotoxin by rumen microbes. For example, mycotoxin DON was almost completely bio-transformed to de-epoxy DON (94-99%) an non-toxic metabolite by rumen microbes (Seeling et al., 2006). But despite the noted rumen mycotoxin degradation, it was noted that ruminants are not completely exempt from toxic effects of mycotoxins as evidenced by deaths regularly reported in ruminants caused by mycotoxin contamination (Marczuk et al., 2012). This raises specific concerns for the presence of noticeable toxic effects in ruminants despite primary considerations that ruminants are less susceptible to mycotoxins due to degradation of mycotoxin by rumen microbes. The current article thus focuses on providing the plausible mechanistic explanation for the occurrence of toxic effects of mycotoxins in ruminants. The article initially lists the mycotoxin toxic effects despite lack of evident clinical manifestation. Then the article

explains how the mycotoxin occurrence factors and rumen microbial factors are responsible for the reported mycotoxin toxic effects in ruminants.

Adverse Effect of Mycotoxins in Ruminants Despite Degradation by Rumen Bacteria

The toxic effects of mycotoxins in ruminants are noticed as deleterious effects on animal production as well as on health. The detailed toxic effects of mycotoxins in ruminants have been saliently described elsewhere by other authors (Gallo et al., 2015; Iheshiulor et al., 2011 and Ogunade et al., 2018). The current article rather focuses on the question that 'why is the presence of toxic effects of mycotoxins in ruminants despite microbial degradation and more importantly despite the mycotoxin level in feed being within permissible regulatory limits?'. Indeed, various factors influence the outcome of the toxic effects of mycotoxins that involves a complex interaction of animal-related factors with dietary changes based on the specific age-needs, toxin-related factors and the envirnmental /manage-mental factors as summarised by Gallo et al. (2015). Briefly, the presence of mycotoxins could a) affect the production and performance of animals, despite lack of toxic effect; b) exert sub-clinical toxic effect despite absence of clinical toxicity; or c) cause clear toxic effect. These three different outcomes are explained below.

While mortality is the easiest identifiable toxic effect, despite reports of deaths from mycotoxins mortality is not always the manifested clinical sign of mycotoxicosis. More frequently mycotoxicosis in ruminants occurs as a range of non-specific symptoms negative impact on animal health, productivity and performance (Kiyothong et al., 2012). Reduced rumen motility (AFB1) (Cook et al., 1986), reduced microbial protein synthesis (DON) (Danicke, 2002), switch in the rumen microbial community composition (DON) (Seeling et al., 2006) to inhibition the multiplication of cellulolytic rumen microorganisms (fusaric acid) (May et al., 2000) are some of the general effects of presence of mycotoxins in ruminant feed, noticed even in absence of evident toxicity. This further explains the negative effect of mycotoxins on

feed intake and feed conversion efficacy, that has been thoroughly discussed by Rodrigues (2014), in-turn affecting animal performance. Further, in situation where the mycotoxins exert sub-clinical toxic effect without apparent the clinical symptomatic disease the negative impact on animal health, productivity and performance is more pronounced, compared to their effect on animal performance without any toxicity. This is more so, in ruminants. Due to the rumen degradation of mycotoxins, the level of exposure is considered generally low, however chronic low dose exposure leads to sub-clinical effects that are evidenced to a higher degree in terms of reduction in performance trait (Whitlow and Hagler, 2005). Reduced milk yield, reproductive performance, higher disease incidence (Whitlow and Hagler, 2005) are all some of the common effects of mycotoxicosis in ruminants. Increased somatic cell count (SSC), loss of body condition, lameness with no apparent disease such as reported cases of increase in mastitis and laminitis, correlated to DON exposure in Nothern-Europe and increase in lameness (subclinical laminitis) and impaired fertility (cystic ovaries) in dairy herds following AFB exposure have been reviewed by Rodrigues (2014). While effects such as increased SCC reduce farm profitability, other immune effects such as increased incidence of disease and the resultant increased animal-to-human transmission of pathogens are of public health concern (Oswald et al., 2005). Indeed studying immune effects is crucial, but of all the parameters assessed, immune-toxicity of mycotoxins in ruminants has been given less priority as compared with other farm animals and rodents. It is imperative to focus on immune effect, as impaired immune response to mycotoxin exposure has been demonstrated to occur at levels that had no effect on growth rate in other species (Smith and Hamilton, 1970). Furthermore, ruminants were demonstrated to be more susceptible to the immunotoxic effects of mycotoxins than pig and poultry immune cells, which occur at low doses corresponding to the low dose long term exposure. However, many immune effects including evaluation of vaccine response, outcome of a disease due to interaction of mycotoxin in the pathogenicity of pathogens requires to be elucidated for many mycotoxins in ruminants. Additionally, many deleterious effects were seen in cattle even when feed mycotoxin levels were within regulatory limits. Such as reduced IgA concentrations (Korosteleva et al., 2007), depressed neutrophil phagocytic activity in cows fed DON contaminated diet 3.5mg/kg dry matter (Korosteleva et al., 2009) and low dose DON (0.6mg/kg of dry matter) long term (63 days) exposure in dairy cattle showed negative influence on somatic cell count, blood parameters and immunity (Jovaisiene et al., 2016). Most of these toxic effects were associated with immuno-toxicity. More importantly, recent research results show shocking results that mycotoxin metabolites of microbial degradation that were previously considered non-toxic (Example: de-epoxy DON metabolite of microbial degradation of DON) is found to be toxic to bovine ovarian cells (Guerrero-Netro et al., 2017 and Pizzo et al., 2016). These data suggest that ruminants are more-or-less at the same level of risks from mycotoxins as monogastric animals such as pigs, dogs, etc. This makes studies on mycotoxin-toxicity in ruminants a necessity than an option, for mitigating the mycotoxin problem and ensuring global public health.

Factors that Complicate the Exposure of Ruminants to Mycotoxins

Indeed the different sources of ruminant diet complicate the actual exposure of ruminants to mycotoxins. Ruminants diets are composed of forages (that includes fresh grass, hay, straw and/or silages) and concentrates (that includes cereals and/or protein feed - such as oil seed cakes) (Gallo et al., 2015). Due to this considerable diversity of the feeding systems adopted and the diverse feeds used in their diets, ruminants in particular are exposed to higher variety of mycotoxins coming both from field mycotoxins (occurs before the harvest - forage, cereals, silage) and storage mycotoxins (occurs after the harvest - cereals, concentrates, silage making) (Gallo et al., 2015). Forages make up a major portion of the ruminant diet and could be contaminated by the mycotoxins in the field as well during storage, resulting from poor storage management practices.

Presence of mycotoxins in straw and silages is also fairly common as evident by surveys (Biomin, 2016). Trichothecenes (DON, T-2), zearalenone, fumonisins, aflatoxins and ergot alkaloid are some of the field-derived mycotoxins, in forages and cereals used for ruminant diets (Driehuis, 2013). Silage in particular, could have mycotoxins from pre-harvest, while ensiling and after ensiling. Roquefortine C, mycophenolic acid, gliotoxin, fumigaclavines, monacolin K and citrinin are the mycotoxins that occur during ensiling process (Driehuis, 2013). The presence of mycotoxins in silage is extensively reviewed in other articles (Ogunade et al., 2018; Wambacq et al., 2016) and could be referred for more detailed description. Further, cereals that make up the major portion of concentrates added to enhance the energy density in ruminant feed are known to be commonly contaminated by mycotoxins. In a 3 year survey Rodrigues and Naehrer (2012), presented evidence that 81 per cent of livestock feed (cereals, distillery grains, finished feed) samples collected worldwide were found to contain at least one mycotoxin. Moreover, a study reported two or more mycotoxins in 48 per cent of the samples. Similarly, contamination of cereals, silage and complete feed were reported in a 4 year survey in Poland (Kosicki et al., 2016).

Additionally, various factors further influence the risk of higher mycotoxin 'exposure and load' in ruminants compared to monogastric animal. As an example based on report by Mostrom and Jacobsen (2011), in the United States during 2008, approximately 25 to 30 per cent of the agricultural commodities were retained for on-farm use and farms where the forage is farm-grown and silage is farm-made, the chances of both field and storage mycotoxin production is higher in ruminant feed (Mostrom and Jacobsen, 2011). Further, the increased need to feed nutrient dense diets to dairy cattle to meet their greater nutrition demand required to accommodate for the higher milk production, increases their chances to mycotoxin exposure from concentrates. Adding to this, ruminants are at higher risk for mycotoxin exposure also due to common consideration that ruminants are less susceptible to

mycotoxin in feed on-account of mycotoxin detoxification by rumen microbes. This conception leads to common practice of diverting the spoiled feed of more susceptible species such as pigs to cattle feeding (Mostrom, 2022).

Rumen Microbiota Plays a Key Role in Toxicity of Mycotoxin in Ruminants

Indeed, it has been demonstrated that some of the common mycotoxins found in animal feed (including DON and OTA) are degraded in the rumen, thereby enhancing the resistance of ruminants to feed borne mycotoxins. However, tolerance of ruminants to mycotoxins is a 'consideration' and not a clear-cut conclusive reality (Fink-Gremmels, 2008). For example, 'high-yielding cows fed energy rich diets', as well as 'young ruminants' and 'animals in the transition period' are more susceptible to the negative effects of these mycotoxin toxic metabolites (Rodrigues, 2014). In addition, species difference also plays an intricate role in symptoms of mycotoxicosis. For example, sheep are more tolerant to roquefortine C (Ogunade et al., 2018), while more susceptible to zearalenone (Knutsen et al., 2017), than cattle. Thus, as stated by Rodrigues (2014), ruminal metabolization of mycotoxins is not equivalent to complete toxininactivation. Evidently, rumen microbial community carries out this ruminal mycotoxin degradation. Thus the actual extent of ruminal mycotoxin degradation and absorption is dependent on at-least three factors: a) rumen microbial population, b) feeding regime and c) the anti-microbial properties of the mycotoxins themselves. The details of this multi-factor dependence are explained below.

Rumen Microbial Population

While rumen detoxification is true for certain mycotoxins (such as DON), but it is not a fool-proof process that ensures complete detoxification. As the ruminal degradation of mycotoxins is rumen microbiota dependent, factors that influence the stability/activity/diversity of rumen microbiota such as dietary composition and the resulting alterations in pH, could impact the mycotoxin degradation potential. It has been shown that depending on

various factors such as dietary starch intake, rumen pH, microbial activity, level of mycotoxin, etc. The degradation and in-turn bioavailability of mycotoxins varies substantially. Bioavailability of AFB1 and OTA were increased in starch-induced low ruminal pH in non-lactating dairy cows (Pantaya et al., 2014, 2016). Further, not all mycotoxins are degraded in rumen (fuminosins, aflatoxins) and on the other-hand certain mycotoxins are transformed into more toxic metabolite (zearalenone to α-zearalenol) (Danicke et al., 2005). Globally, dairy cattle under the conditions of modern feeding strategies fed diets containing high-energy low fibre are more prone to sub-acute ruminal acidosis (SARA). SARA combined with increased stress of high performance and compromised microbial activity could lead to more mycotoxins escape rumen detoxification and in-turn more mycotoxins absorbed in intestines.

More importantly, differences in mycotoxin degradation between young and adult ruminants enhance the susceptibility of young ruminants to mycotoxicosis. Although calves carry the same diversity of microbiota in their rumen as adult cattle, the proportions are not the same (Jami et al., 2013). The condition could be explained by the fact that although, weaned calves are technically capable of surviving on a complete solid feed based diet, including forages, their diet is mostly concentrate based. Further, they are not completely capable of deriving their nutritional requirement completely from forages until around 4-5 months. Yet, it takes almost 6 months for their rumen to resemble the adult microbial ratios (Jami et al., 2013) and 2 years to be completely similar (Dill-Mcfarland et al., 2017). Thus, emphasizing that ruminants are not always resistant to mycotoxin toxicity.

Feeding Regime

Feeding regime has a profound influence on the rumen pH and microbial diversity. It is well known that providing higher concentrates (such as in modern intensive farming system) the rumen pH decreases and the cattle are more prone to sub-acute ruminal acidosis (SARA). However, during ruminal acidosis (excess

energy feed) transformation of mycotoxins in a compromised. In-vitro studies on rumen fluid had shown that starch based diet, which is primary concern for SARA reduces the degradation rate of DON than cellulose diet (Jeong et al., 2010). Continuing with the example of DON, in-vitro studies suggests that its degradation by primary rumen culture was only 35 per cent as opposed to 100 per cent by chicken large intestinal contents (He et al., 1992). Moreover, the biotransformation of DON was inhibited at pH 5.2, driving the authors to suggest that the detoxification of DON is pH dependent. Moreover, despite a faster transformation of DON into DOM-1 by the original rumen culture, the proportion of detoxification was reduced for subsequent subcultures (He et al., 1992).

Incidentally in-vivo experiments show that bioavailability of certain mycotoxins (AFB1, OTA) increases in starch-induced low ruminal pH (Pantaya et al., 2016). Adding to this, under conditions of high concentrate diet that favour ruminal acidosis or SARA, there is alteration in the microbial activity or diversity in rumen (AlZahal et al., 2017) as well as an evident increased compromise of rumen mucosal barrier function (Owens et al., 1998). Such altered microbial activity (with or without altered rumen mucosal barrier function) could be the possible cause for the compromised detoxification and increased bioavailability of mycotoxins as demonstrated by Pantaya et al. (2016). Particularly rumen pH of calves is mostly acidic around weaning up-to 12 weeks of age (Quigley et al., 1992 and Suarez-Mena et al., 2015), thus calves are more prone to SARA. This raises the risk of calves to exposed mycotoxins, due to increased absorption.

The Anti-Microbial Properties of the Mycotoxins

Mycotoxins are known for their antibacterial properties. Most of the mycotoxins common in animal feed including DON, FB1, T-2, AFB1 have antibacterial effect (Ali-Vehmas *et al.*, 1998 and Bisht *et al.*, 2011). Subsequently, mycotoxins have been demonstrated to negatively impact certain aspects of rumen fermentative capacity both *in-vitro* and *in-vivo*.

As an example, DON was shown to reduce ammonia-N and total gas production, *in-vitro* (Jeong *et al.*, 2010), whereas in adult cattle, the reduced efficiency of microbial protein synthesis (24%) in rumen and the flow of microbial protein into the duodenum (21%) (Danicke *et al.*, 2005). Such effects on nutrient utilization could be more pronounced in calves and other young ruminants due to still developing rumen and rumen microflora.

Overall, three different types of factors that include a) mycotoxin factor (mycotoxin dose and the anti-microbial activity of mycotoxins); b) animal factor (age group of animal, the rumen microbial population and the rumen digestion kinetics), as well as c) feed related factor (feed composition, more specifically relating to starch and the fibre content in the diet) are the key factors that decide the actual rate of degradation/absorption of mycotoxins in ruminants. This in-turn decides the resultant toxic effects noticed in ruminants, despite the initial consideration of less susceptibility of ruminants to mycotoxins due to rumen microbial mycotoxin-metabolization. While it seems that generally young ruminants are more susceptible to mycotoxicosis due to lack of ability to efficiently degrade mycotoxins than adults. Yet, susceptibility of adult ruminants to mycotoxins could not be neglected.

Studies of mycotoxins in ruminants have received less attention than in monogastric farm animals (pigs and poultry). But, despite previous consideration that ruminants are less susceptible to mycotoxins due to rumen microbial degradation, the extent of mycotoxindetoxification depends on the mycotoxin-dose, animal age group, feeding regime, rumen digestion dynamics and the microbial community, thus increasing susceptibility of ruminants of different age groups to mycotoxins. Further mycotoxins-metabolites of microbial degradation (de-epoxy-DON), previously considered non-toxic was shown to be toxic by recent research in cattle. Additionally, some of the mycotoxin-metabolites produced by rumen microbial degradation are more toxic than parent compound (Zearalenone to α -Zearalenol). As evident, for most of the known mycotoxins either their complete toxic profile/mechanism of action/toxic dose are not completely available, in ruminants. In this regard, with view of growing risk of mycotoxin contamination of ruminant feed, it is imperative to screen the mycotoxins for their toxicity in ruminants relying on more and more sensitive markers.

REFERENCES

- ALI-VEHMAS, T., RIZZO, A., WESTERMARCK, T. AND ATROSHI F., 1998, Measurement of antibacterial activities of T-2 toxin, deoxynivalenol, ochratoxin A, matoxin B1 and fumonisin B1 using Microtitration tray-based turbidimetric techniques. *J. Vet. Med.*, **45** (1): 453 458.
- ALZAHAL, O., LI, F., GUAN, L. L., WALKER, N. D. AND MCBRIDE, B. W., 2017, Factors influencing ruminal bacterial community diversity and composition and microbial fibrolytic enzyme abundance in lactating dairy cows with a focus on the role of active dry yeast. *J. Dairy. Sci.*, **100** (6): 4377 4393.
- BATTILANI, P., TOSCANO, P., VAN DER FELS-KLERX, H. J., MORETTI, A., CAMARDO LEGGIERI, M., BRERA, C., RORTAIS, A., GOUMPERIS, T. AND ROBINSON, T., 2016, Aflatoxin B1 contamination in maize in Europe increases due to climate change. *Sci. Rep.*, 6 (3):1-7.
- BIOMIN, 2016, World mycotoxin survey. biomin feed surv., Annual Reports, Inzersdorf-Getzersdorf, pp.: 1-10.
- BIOMIN, 2020, World mycotoxin survey. Biomin feed surv., Annual Reports, Inzersdorf-Getzersdorf, pp.: 1 10.
- BISHT, S. S., PRAVEEN, B., PANDA, A., BEHERA, S., PANDA, K. K., MISHRA, R. AND PATRO, S. K., 2011, Comparative study of various mycotoxins against few bacterial test organisms. *Int. J. Pharm. Pharm. Sci.*, 3 (5): 288 291.
- Cook, W., Richard, J., Osweiler, G. and Trampel, D., 1986, Clinical and pathologic changes in acute bovine aflatoxicosis/: Rumen motility and tissue and fluid concentrations of aflatoxins. *Ame. J. Vet. Res.*, **47** (8): 1817 - 1825.

- Danicke, S., 2002, Effects of fusarium toxin contaminated wheat grain and of a detoxifying agent on rumen physiological parameters and in sacco dry matter degradation of wheat straw and lucerne hay in wethers. J. Anim. Feed. Sci., 11 (3): 437 - 451.
- Danicke, S., Matthaus, K., Lebzien, P., Valenta, H., Stemme, K., Ueberschar, K. H., Razzazi-Fazeli, E., Bohm, J. and Flachowsky, G., 2005, Effects of fusarium toxin-contaminated wheat grain on nutrient turnover, microbial protein synthesis and metabolism of deoxynivalenol and zearalenone in the rumen of dairy cows. *J. Anim. Physiol. Anim. Nutr. (Berl)*, **89** (9-10): 303 315.
- DILL-MCFARLAND, K. A., BREAKER, J. D. AND SUEN, G., 2017, Microbial succession in the gastro intestinal tract of dairy cows from 2 weeks to first lactation. *Sci. Rep.*, 7 (16): 1 - 12.
- Driehuis, F., 2013., Silage and the safety and quality of dairy foods/: A review contamination pathway from silage to raw milk. *Agri. Food. Sci.*, **22** (1):16 34.
- FDA, 2021, Certain lots of sportmix pet food eecalled for potentially fatal levels of aflatoxin. FDA alerts., FDA Outbreaks and Advisories, Silver Spring, pp.: 1 3.
- FINK-GREMMELS, J., 2008, The role of mycotoxins in the health and performance of dairy cows. *Vet. J.*, **176** (1): 84 92.
- Gallo, A., Giuberti, G., Frisvad, J. C., Bertuzzi, T. and Nielsen, K. F., 2015, Review on mycotoxin issues in ruminants: Occurrence in forages, effects of mycotoxin ingestion on health status and animal performance and practical strategies to counteract their negative effects. *Toxins.*, 7 (8): 3057 3111.
- Grenier, B. and Oswald, I., 2011, Mycotoxin co-contamination of food and feed: meta-analysis of publications describing toxicological interactions. *World Mycotoxin J.*, 4 (3): 285 313.
- Guerrero-Netro, H. M., Estienne, A., Chorfi, Y. and Price, C. A., 2017, The mycotoxin metabolite deepoxy-deoxynivalenol increases apoptosis and decreases steroidogenesis in bovine ovarian theca cells. *Biol. Reprod.*, **97** (5): 746 757.

- HE, P., YOUNG, L.G. AND FORSBERG, C., 1992, Microbial transformation of deoxynivalenol (Vomitoxin). *Appl. Environ. Microbiol.*, **58** (12): 3857 3863.
- IHESHIULOR, O. O. M., ESONU, B. O., CHUWUKA, O. K., OMEDE, A. A., OKOLI, I. C. AND OGBUEWU, I. P., 2011, Effects of mycotoxins in animal nutrition: A review. *Asian J. Anim. Sci.*, 5 (1): 19 33.
- Jami, E., Israel, A., Kotser, A. and Mizrahi, I., 2013, Exploring the bovine rumen bacterial community from birth to adulthood. *ISME J.*, 7 (6): 1069 1079.
- JEONG, J. S., LEE, J. H., SIMIZU, Y., TAZAKI, H., ITABASHI, H. AND KIMURA, N., 2010, Effects of the fusarium mycotoxin deoxynivalenol on *in vitro* rumen fermentation. *Anim. Feed Sci. Technol.*, **162** (3): 144-148.
- JOVAISIENE, J., BAKUTIS, B., BALIUKONIENE, V. AND GERULIS, G., 2016, Fusarium and aspergillus mycotoxins effects on dairy cow health, performance and the efficacy of anti-mycotoxin additive. *Pol. J. Vet. Sci.*, **9** (1): 79 87.
- KIYOTHONG, K., ROWLINSON, P., WANAPAT, M. AND KHAMPA, S., 2012, Effect of mycotoxin deactivator product supplementation on dairy cows. *Anim. Prod. Sci.*, **52** (9): 832 841.
- Knutsen, H. K., Alexander, J., Barregård, L., Bignami, M., Bruschweiler, B., Ceccatelli, S. and Oswald, I., 2017, Risks for animal health related to the presence of zearalenone and its modified forms in feed. *EFSA J.*, **15** (7): 1 123.
- Korosteleva, S. N., Smith, T. K. and Boermans, H. J., 2009, Effects of feed naturally contaminated with fusarium mycotoxins on metabolism and immunity of dairy cows. *J. Dairy Sci.*, **92** (4): 1585 1593.
- Korosteleva, S. N., Smith, T. K. and Boermans, H. J., 2007, Effects of feedborne fusarium mycotoxins on the performance, metabolism and immunity of dairy cows. *J. Dairy Sci.*, **90** (8): 3867 - 3873.
- Kosicki, R., Grajewski, J. and Twaru, M., 2016, Multiannual mycotoxin survey in feed materials and feedingstuffs. *Ani. Feed Sci. Tech.*, **215** (2016): 165-180.

- Lewis, L., Onsongo, M., NJapau, H., Schurz-Rogers, H., Luber, G., Kieszak, S., Nyamongo, J., Backer, L. D., Abdikher, M., Misore, A., Decock, K. and Rubin, C., 2005, Aflatoxin contamination of commercial maize products during an outbreak of acute aflatoxicosis in eastern and central Kenya. *Environ. Health Perspect.*, 113 (12): 1763 1767.
- MAY, H. D., Wu, Q. AND BLAKE, C. K., 2000, Effects of the *Fusarium* spp. mycotoxins fusaric acid and deoxynivalenol on the growth of Ruminococcus albus and Methanobrevibacter ruminantium. *Canadian J. Microbiol.*, **46** (8): 692 699.
- Mostrom, M. S. and Jacobsen, B. J., 2011, Ruminant mycotoxicosis. *Vet. Clin. NA. Food. Anim. Pract.*, 27 (2): 315 344.
- Mostrom, S. M., 2022, Trichothecene toxicosis in animals toxicology MSD veterinary manual. MSD Web Manual, pp. : 1.
- OGUNADE, I. M., DROUIN, P. AND ADESOGAN, A. T., 2018, Silage review: Mycotoxins in silage: Occurrence, effects, prevention and mitigation. *J. Dairy Sci.*, **101** (5): 4034 4059.
- OSWALD, I. P., MARIN, D. E., BOUHET, S., PINTON, P., TARANU, I. AND ACCENSI, F., 2005, Immuno toxicological risk of mycotoxins for domestic animals. *Food Addit. Contam.*, **22** (4): 354 360.
- Owens, F. N., Secrist, D. S., Hill, W. J. and Gill, D. R., 1998, Acidosis in cattle: A review. *J. Ani. Sci.*, **76** (1): 275 286.
- Pantaya, D., Morgavi, D. P., Silberberg, M., Chaucheyrasdurand, F., Martin, C., Suryahadi-Wiryawan, K. G. and Boudra, H., 2016, Bioavailability of aflatoxin B1 and ochratoxin A, but not fumonisin B1 or deoxynivalenol, is increased in starch-induced low ruminal pH in nonlactating dairy cows. *J. Dairy Sci.*, **99** (12): 9759 - 9767.
- Pantaya, D., Morgavi, D. P., Silberberg, M., Martin, C., Suryahadi-wiryawan, K. G. and Boudra, H., 2014, Low pH enhances rumen absorption of aflatoxin B1 and ochratoxin A in sheep. *Glob. Vet.*, **12** (2): 227 232.

- Pizzo, F., Caloni, F., Schreiber, N. B., Cortinovis, C. and Spicer, L. J., 2016, *In vitro* effects of deoxynivalenol and zearalenone major metabolites alone and combined, on cell proliferation, steroid production and gene expression in bovine small-follicle granulosa cells. *Toxicon.*, **109** (2016): 70 83.
- QUIGLEY, J. D., STEEN, T. M. AND BOEHMS, S. I., 1992, Postprandial changes of selected blood and ruminal metabolites in ruminating calves fed diets with or without hay. *J. Dairy Sci.*, **75** (1): 228 235.
- Robbins, C. A., Swenson, L. J., Nealley, M. L., Kelman, B. J. and Gots, R. E., 2000, Health effects of mycotoxins in indoor air: A critical review. *Appl. Occup. Environ. Hyg.*, **15** (10): 773 784.
- Rodrigues, I., 2014, A review on the effects of mycotoxins in dairy ruminants. *Ani. Prod. Sci.*, **54** (9): 1155 1165.
- Rodrigues, I. and Naehrer, K., 2012, A three-year survey on the worldwide occurrence of mycotoxins in feedstuffs and feed. *Toxins* (*Basel*)., 4 (9): 663 675.
- SEELING, K., DANICKE, S., VALENTA, H., VAN EGMOND, H. P., SCHOTHORST, R. C., JEKEL, A. A., LEBZIEN, P., SCHOLLENBERGER, M., RAZZAZI-FAZELI, E. AND FLACHOWSKY, G., 2006, Effects of Fusarium toxin-contaminated wheat and feed intake level on the biotransformation and carry-over of deoxynivalenol in dairy cows. Food Addit. Contam., 23 (10):1008-1020.
- Shephard, G., Berthiller, F., Burdaspal, P., Crews, C., Jonker, M., Krska, R., Macdonald, S., Malone, R., Maragos, C., Sabino, M., Solfrizzo, M., Van Egmond, H. and Whitaker, T., 2012, Developments in mycotoxin analysis: An update for 2010-2011. *World Mycotoxin J.*, **5** (1): 3 30.
- SMITH, J. W. AND HAMILTON, P. B., 1970, Aflatoxicosis in the broiler chicken. *Poult. Sci.*, **49** (1): 207 215.
- Streit, E., Naehrer, K., Rodrigues, I. and Schatzmayr, G., 2013, Mycotoxin occurrence in feed and feed raw materials worldwide: Long-term analysis with special focus on Europe and Asia. *J., Sci., Food Agric.*, **93** (12): 2892 2899.

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- SUAREZ-MENA, F. X., HEINRICHS, A. J., JONES, C. M., HILL, T. M. AND QUIGLEY, J. D., 2015, Digestive development in neonatal dairy calves with either whole or ground oats in the calf starter. *J. Dairy Sci.*, **98** (5): 3417 3431.
- Susan, S. L., 2006, Dogs keep dying: Too many owners remain unaware of toxic dog food. Cornell University Chronicle. New York, pp.: 1.
- Wambacq, E., Vanhoutte, I., Audenaert, K. and Gelder, L., D., 2016, Occurrence, prevention and remediation of toxigenic fungi and mycotoxins in silage/: A review. *J. Sci. Food Agric.*, **96** (7): 2284 2302.
- WHITLOW, L. AND HAGLER, W. M., 2005, Mycotoxins in dairy cattle: Occurrence, toxicity, prevention and treatment. *Proc. Southwest Nutr. Conf. Arizona*, pp.: 124 - 138.
- WHITLOW, L. W. AND HAGLER, W. M., 2008, Mold and mycotoxin issues in dairy cattle: Effects, prevention and treatment, WCDS Advances in Dairy Technology, Red Deer, Alberta, pp.: 195 209.